

On page 18 please replace lines 18-22 with the following:

*B 2*  
--- A clinical isolate of the *Candida albicans*, was obtained from a patient with vaginal thrush. The identity of the *Candida* species was confirmed with the use of an API® 20C Auxonogram strip (API System S.A., France). The *C. albicans* isolate was designated KEMHS. ---

On page 19 please replace lines 3-11 with the following:

*B 3*  
--- *Candida* cells were ruptured mechanically with the use of a DYNOMILL™ (WAB, Switzerland). Milling was continued until 99% cell disruption was obtained. The soluble *Candida* cell extracts were collected and dispensed into 50ml centrifuge tubes. The extracts were centrifuged for 12h at 8,517 x g and 4°C to precipitate insoluble cell walls. The supernatants containing the soluble cytoplasmic antigen fraction were recovered and passed through a 0.45µm filter membrane.

Please replace the paragraph bridging pages 19 and 20 with the following:

*B 4*  
--- The soluble cytoplasmic antigen fraction was dialysed overnight against 20mM Tris.Cl, pH7.4. An estimate of the quantity of protein in solution was performed using the BIO-RAD® (Bradford) microassay procedure in accordance with the manufacturers instructions. A portion of the cytoplasmic antigen extract was analysed by SDS-PAGE. ---

On page 20 please replace lines 10-26 with the following:

*B 5*  
--- Purification of the enolase antigen was conducted in the same fashion as the soluble *Candida* cytoplasmic antigen except that it was not subjected to Con A-Sepharose chromatography. Instead, following dialysis and filtering through a 0.20µm syringe filter (cellulose acetate), the filtered extracts were applied to a Pharmacia Biotech XK 50/20 chromatography column packed with Pharmacia Biotech Source 15Q quaternary ammonium anion exchanger (Pharmacia LKB, Uppsala, Sweden). The column was equilibrated prior to chromatography with column binding buffer 'A' (20mM bis-Tris, pH 6.5). Anion exchange chromatography of the crude extracts was controlled and recorded using the BIO-RAD® and

ECONO® system (Bio-Rad Laboratories, USA). Bound protein was eluted from the column with a salt gradient of buffer 'B' (1M NaCl in buffer 'A', pH 6.5). The recovered fractionated proteins were analysed by an enzyme activity assay. - - -

Please replace the paragraph bridging pages 20 and 21 with the following:

36  
--- The active enzyme enolase hydrolyses D(+)2-phosphoglyceric acid (PGA) to phosphoenolpyruvate (PEP). The production of PEP can be monitored by spectrophotometry at 240nm. 20 $\mu$ l of protein solution was combined with 1ml of enolase substrate solution (50mM Tris-HCl pH 7.4, 2.7mM magnesium acetate, 1.0mM EDTA, 1.2mM D(+)2-phosphoglyceric acid) in a quartz cuvette and the change of absorbance recorded at 1min intervals. The specific activity was defined as the conversion of 1 $\mu$ mol of PGA to PEP per mm per mg protein. An estimate of the quantity of protein in solution was performed using the BIO-RAD® (Bradford) microassay procedure. - - -

2-16  
On page 21 please replace lines 10-26 with the following:

37  
--- Eluate fractions containing enolase activity were selected and dialysed for 12h at 25°C in h<sub>2</sub>O. The dialysed fractions were recovered and filtered through a 0.20 $\mu$ m syringe filter. The filtrate was concentrated ten-fold by evaporation under vacuum for 5h. The concentrated samples were dialysed with binding buffer 'A' (10mM sodium acetate, pH 4.7) immediately prior to application to a Pharmacia Biotech Mono S HR10/10 chromatography column packed with methyl sulphonate cation exchanger (Pharmacia LKB, Uppsala, Sweden). Cation exchange chromatography performed using the BIO-RAD® Biologic system. Bound protein fractions were eluted from the column with a salt gradient of buffer 'B' (1M NaCl in buffer 'A', pH 4.7). Fractions containing enolase activity were identified by the enzyme activity assay described above. - - -

At the end of the specification, after the claims, please insert the page titled "ABSTRACT OF THE DISCLOSURE."

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B8  
002.1002872.2  
Abstract  
--- The present invention relates to a method and a means of diagnosing *Candida* infection. In particular, the present invention relates to a method of diagnosing *Candida*